

## Rekom Syphilis High-Quality Raw Material for 3rd generation ELISA assay

NAME	CAT NUMBER	SOURCE	APPLICATION	DESCRIPTION
TmpA	RAG0073	E. coli	WB, DB, IE, DE, CLIA, LF	Membrane lipoprotein
Tpp15	RAG0009	E. coli	WB, DB, IE, DE, CLIA, LF	Membrane lipoprotein
	RAG0009BIOT	E. coli	WB, DB, CE, DAS, NP, PO	Tpp15 biotinylated
Tpp17	RAG0008	E. coli	WB, DB, IE, DE, CLIA, LF	Membrane lipoprotein
	RAG0008BIOT	E. coli	WB, DB, CE, DAS, NP, PO	Tpp17 biotinylated
Tpp47	RAG0010	E. coli	WB, DB, IE, DE, CLIA, LF	Membrane lipoprotein
	RAG0010BIOT	E. coli	WB, DB, CE, DAS, NP, PO	Tpp47 biotinylated
ChimSyphilis1	RAG0046 🥋	E. coli	WB, DB, IE, DE, CLIA, LF	Recombinant chimeric antigen (Tpp17 and Tpp47)
	RAG0046BIOT	E. coli	WB, DB, CE, DAS, NP, PO	ChimSyphilis1 biotinylated
ChimSyphilis2	RAG0064	E. coli	WB, DB, IE, DE, CLIA, LF	Recombinant chimeric antigen (Tpp15 and TmpA)
	RAG0064BIOT	E. coli	WB, DB, CE, DAS, NP, PO	ChimSyphilis2 biotinylated

Coloured text boxes show matched raw material for double antigen sandwich (DAS) systems: the non-biotinylated protein should be used as a capturer, coating the plate and the mono or poly-biotinylated as a detector of the system.

WB: Western Blot DB: Dot Blot IE: Indirect ELISA

DE: positive control in direct ELISA
CLIA: Chemiluminescent Immunoassay

LF: Lateral Flow
CE: Capture ELISA
DAS: Double antigen sar

DAS: Double antigen sandwich NP: nanoparticles binding PO: plate orientation

Pack size: 0.1 mg\*; 1 mg; bulk Format: liquid; lyophilised

\*under availability, for liquid format

8

Top product (Satisfaction guarantee)

## PRODUCT PERFORMANCE:

- Versatility
- Validation
- ▶ Conjugation
- ▶ Reproducibility
- Broad spectrum
- Specificity and sensitivity
- Technical support
- Costs reduction and fast delivery



Syphilis is a multistage progressive disease caused by the spirochete *Treponema pallidum* subsp. *pallidum* and is characterized by localised, disseminated and chronic stages. Manifestations include the development of a localised lesion called a chancre during the primary stage and disseminated skin

lesions and meningovascular syphilis during the secondary stage, followed by a period of latency lasting from months to decades. Since direct microscopy is possible only when lesions are present, and this is not the case in the majority of patients, detection of antibodies against *T. pallidum* is the most effective method for the diagnosis of syphilis.

Enzyme immunoassays have shown some advantages in relation to the tests used for the laboratory diagnosis of syphilis since they are easy and quick to perform and objective to read. They also have the potential to be automated.

The diagnosis of congenital syphilis, although useful in documenting maternal infection, does not distinguish maternal from infant antibody. Serologic tests detecting IgM antibodies to infectious agents are useful in diagnosing congenital infections. Since IgA antibodies, like IgM, do not cross the placenta, they are also potential markers of congenital infection. A consistent finding is the IgM and IgA reactivity to the 47-kDa antigen by sera from infants at risk for congenital syphilis.

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