

## Recombinant antigen p30 (SAG1) for *Toxoplasma gondii*

CATALOG NUMBER: RAG0011

LOT NUMBER: #

**RECOMBINANT ANTIGEN:** *T. gondii* antigen p30 (Santoro et al., 1986).

**DESCRIPTION:** the *T. gondii* antigen p30 has been prepared as a recombinant antigen fused to a his-tag in its N-terminal. It corresponds to the *T. gondii* major surface antigen (SAG1).

**PRESENTATION:** liquid protein solution

**SOURCE:** *Escherichia coli*

**MOLECULAR WEIGHT:** determined by SDS-PAGE, the protein band is between molecular markers of 45,000-35,000 Da, while relative molecular mass calculated from amino acid sequence is 36,686.1 Da.

**BATCH COMPOSITION:**

COMPONENTS	COMPOSITION
his-p30	recombinant antigen with a his-tag in its N-terminus
Storage buffer	20 mM phosphate buffer pH 8, 0.15 M NaCl and 0.1% polyoxyethylene (10) tridecyl ether

### QUALITY CONTROL:

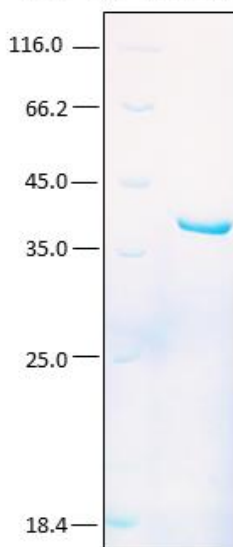
#### 1. PROTEIN CONCENTRATION DETERMINED ESPECTROPHOTOMETRICALLY

DO<sub>280</sub> = 0.761  
 $A_{0.1\%} (=1 \text{ g/l}) = 0.633$   
 CONCENTRATION\*: 1.2 mg/ml

\* The measurement of the protein concentration has been performed with the theoretical extinction coefficient of the recombinant protein obtained from Gill and vonHippel, 1989. It is recommended that the users carry out their absorbance determinations to avoid equipment variabilities regarding final concentration, mainly in reproducibility analysis.

#### 2. PURITY CONTROL IN SDS-PAGE: 15%

kDa Mw p30 (SAG1)

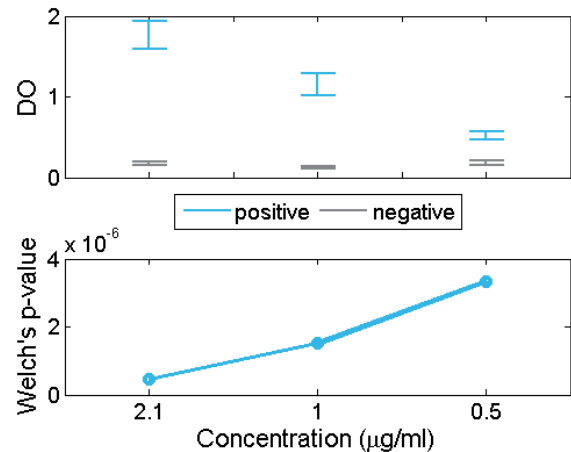


**Figure 1.** SDS-PAGE analysis (15%) of 2 µl of recombinant p30. Purity is >95% as determined by gel electrophoresis.

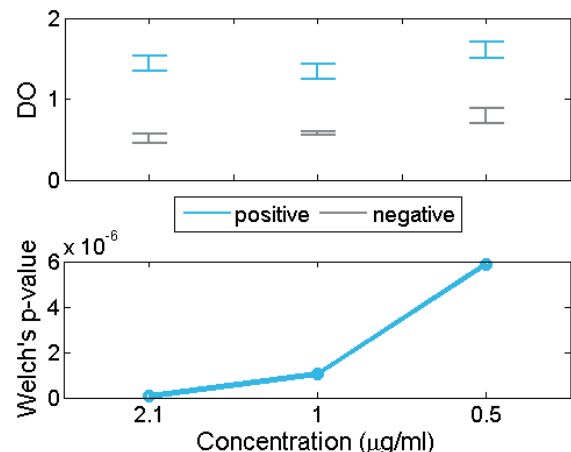
### 3. TITRATION CURVE BY AN ELISA ASSAY

The titer has been suggested in reference to an "in-house" ELISA kit performed at Rekom Biotech over the first lot obtained.

Each end user should carry out his own titration for his particular application.

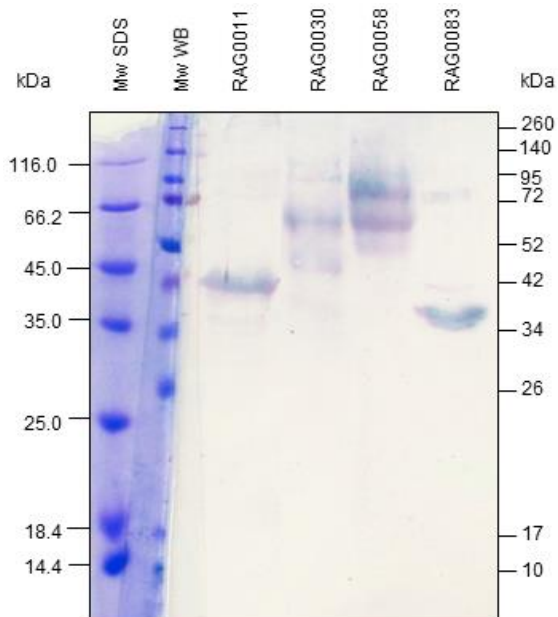


**Figure 2.** In this plot, the optical density at 450/620 nm for positive (blue) and negative (gray) Toxo IgG sera are compared for each concentration of the recombinant antigen. An appropriate statistical test of significance for the comparison of means between both groups, the Welch's test, is employed. Eligible concentrations for the use of the antigen should present statistically significant differences between positive and negative sera. This happens when the intervals at the top do not overlap and, equivalently, when the p-value at the bottom is below 0.05. In the present figure, all p-values are below 0.05 and thus the intervals do not overlap. Therefore, any of the showed concentrations can be used to distinguish between positive and negative sera.



**Figure 3.** In this plot, the optical density at 450/620 nm for positive (blue) and negative (gray) Toxo IgM sera are compared for each concentration of the recombinant antigen. An appropriate statistical test of significance for the comparison of means between both groups, the Welch's test, is employed. Eligible concentrations for the use of the antigen should present statistically significant differences between positive and negative sera. This happens when the intervals at the top do not overlap and, equivalently, when the p-value at the bottom is below 0.05. In the present figure, all p-values are below 0.05 and thus the intervals do not overlap. Therefore, any of the showed concentrations can be used to distinguish between positive and negative sera.

#### 4. WESTERN BLOT ANALYSIS



**Figure 4.** When a polyclonal Ab obtained by using a clarified native extract of sporozoite oocysts of *Toxoplasma gondii* as immunogen was used in a western blot analysis, clear signals were obtained in our recombinant antigens for *T. gondii* RAG0030, RAG0011, RAG0083 and RAG0058.

#### 5. ABSENCE OF PRECIPITATION AFTER A FREEZING AND THAWING CYCLE: ok

#### LOT SPECIFICATIONS:

1. **CONCENTRATION:** 1.2 mg/ml
2. **TOTAL QUANTITY PER ALIQUOT:** 1 mg
3. **TOTAL VOLUME PER ALIQUOT:** 0.875 ml
4. **SUGGESTED TITER BY ELISA:** up to 2,400, which corresponds to 0.5 µg/ml of protein concentration in plates for IgG and IgM detection. Better P/N ratios in IgG detection are obtained at protein concentrations of 0.11 µg/ml.

**Important Notes:** During shipment, small volumes of product will occasionally become entrapped in the seal of the product vial. For products with volumes of 200 µl or less, we recommend gently tapping the vial on a hard surface or briefly centrifuging the vial in a tabletop centrifuge to dislodge any liquid in the containers cap.

Although recombinant antigens are expressed in non-pathogenic *E. coli* and bacterial integrity is destroyed during purification, the antigen preparation should be handled as potentially infectious.

**FOR RESEARCH AND COMMERCIAL USE *IN VITRO*: not for human *in vivo* or therapeutic use.**

**5. STORAGE:** Protein is shipped with dry ice. Upon arrival, it should be aliquoted to avoid repeated freezing and thawing cycles and stored at -20°C to -80°C. In order to defrost the protein, maintain the aliquot at 25°C without shaking to avoid aggregation.

**6. TESTED APPLICATIONS:** ELISA and western blot.

**7. POSSIBLE APPLICATIONS:** WB, DB, Indirect ELISA, positive control in direct ELISA, CLIA, lateral-flow. Where this product has not been tested for use in a particular technique, this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates.

**8. OBSERVATIONS:** proteins should be maintained frozen at high concentrations. The dilution to be performed for ELISA assays should be made with a small quantity of protein, the same day of the experiment. In order to defrost the protein, maintain the aliquot at 25°C without shaking to avoid aggregation. Prior making test dilutions and after defrosting the protein, is recommended to remove possible protein aggregates by centrifuging the stock solution, avoiding alterations in the immobilization of the biomolecule to the solid surface.

#### RELATED PRODUCTS:

p29 (GRA7), p30 (SAG1) (RAG0019, RAG0030), p35 (GRA8), ChimToxo1.

#### BIBLIOGRAPHY:

**Santoro, F., Charif, H. and Capron, A.** The immunodominant epitope of the major membrane tachyzoite protein (p30) of *Toxoplasma gondii*. 1986, *Parasite Immunol.* 8:631-9.

**Gill SC, von Hippel PH.** Calculation of protein extinction coefficients from amino acid sequence data. *Anal Biochem.* 1989 Nov 1;182(2):319-26.